

Mouse IgG1 ELISA Kit

Life Diagnostics, Inc., Catalog Number: 5010-1A

ELISA for the Quantitative Determination of Mouse IgG1 in Serum or Plasma

INTRODUCTION

The mouse IgG1 ELISA kit is designed for measurement of IgG1 in mouse serum or plasma. The assay uses goat anti-mouse IgG for solid phase (microtiter wells) immobilization and horseradish peroxidase (HRP) conjugated goat anti-mouse IgG1 antibodies for detection. The capture antibodies were cross-adsorbed on mouse IgM and IgA agarose columns, and the detection antibodies were cross adsorbed on mouse IgM, IgA, IgG2a, IgG2b and IgG3 agarose thereby ensuring specificity for IgG1. Cross-reactivity with immunoglobulins from other species has not been investigated.

PRINCIPLE OF THE TEST

Test samples are diluted and incubated in the microtiter wells for 45 minutes alongside prepared mouse IgG1 standards. The microtiter wells are subsequently washed and HRP conjugate is added and incubated for 30 minutes. IgG1 molecules are thus sandwiched between the immobilization and detection antibodies. The wells are then washed to remove unbound HRP-labeled antibodies and TMB Reagent is added and incubated for 20 minutes at room temperature. This results in the development of a blue color. Color development is stopped by the addition of Stop Solution, changing the color to yellow, and optical density is measured spectrophotometrically at 450 nm. The concentration of IgG1 is proportional to the optical density of the test sample and is derived from a standard curve.

MATERIALS AND COMPONENTS

Materials provided with the kit:

- Anti mouse IgG coated 96-well plate (12 strips of 8 wells)
- Anti mouse IgG1-HRP Conjugate, 11 ml
- Reference standard (lyophilized)¹
- 20x Wash Solution, 50 ml
- 10x Immunoglobulin Diluent (25 ml)
- TMB Reagent (One-Step) 11 ml
- Stop Solution (1N HCl), 11 ml

Materials required but not provided:

- Precision pipettes and tips
- Distilled or deionized water
- Polypropylene or glass tubes
- Vortex mixer
- Absorbent paper or paper towels
- Micro-Plate incubator/shaker mixing speed of ~150 rpm
- Plate washer
- Plate reader with an optical density range of 0-4 at 450nm
- Graph paper (PC graphing software is optional)

¹ The reference standard consists of lyophilized mouse serum of known IgG1 concentration in a BSA matrix. The IgG1 concentration was determined relative to mouse IgG1 standard obtained from an independent laboratory.

STORAGE OF THE TEST KIT

The test kit will remain stable for six months from the date of purchase provided that the components are stored as described above. The microtiter plate should be kept in a sealed bag with desiccant to minimize exposure to damp air.

GENERAL INSTRUCTIONS

1. Please read and understand the instructions thoroughly before using the kit.
2. All reagents should be allowed to reach room temperature (18-25°C) before use.
3. Optimum results are achieved if, at each step, reagents are pipetted into the wells of the microtiter plate within 5 minutes.

IMMUNOGLOBULIN DILUENT PREPARATION

The diluent is provided as a 10x stock. Prior to use estimate the final volume of diluent required for your assay and dilute one (1) volume of the 10x stock with nine (9) volumes of distilled or deionized water.

WASH SOLUTION PREPARATION

The wash solution is provided as a 20x stock. Prior to use dilute the contents of the bottle (50 ml) with 950 ml of distilled or deionized water.

STANDARD PREPARATION

1. The mouse IgG1 standard is provided as a lyophilized stock. Reconstitute as detailed on the vial label (***the reconstituted standard is stable at 4°C for one week but should be aliquoted and frozen at -20°C after reconstitution if future use is intended***).
2. Label 6 polypropylene or glass tubes as 50, 25, 12.5, 6.25, 3.13, and 1.56 ng/ml.
3. Into the tube labeled 50 ng/ml, pipette the volume of diluent detailed on the IgG1 standard vial label. Then add the indicated volume of IgG1 standard (shown on the IgG1 standard vial label) and mix gently. This provides the 50 ng/ml standard.
4. Dispense 250 µl of diluent into the tubes labeled 25, 12.5, 6.25, 3.13, and 1.56 ng/ml.
5. Prepare a 25 ng/ml standard by diluting and mixing 250 µl of the 50 ng/ml standard with 250 µl of diluent in the tube labeled 25 ng/ml.
6. Similarly prepare the 12.5, 6.25, 3.13, and 1.56 ng/ml standards by serial dilution.

SAMPLE PREPARATION

General Note: Mouse serum IgG1 concentrations vary with species and treatment conditions. In our laboratory we have measured values of 0.25 to 2 mg/ml or greater. We suggest an

initial dilution factor of 100,000 fold using the following procedure for each sample to be tested:

1. Dispense 498 μ l and 798 μ l of 1x diluent into separate tubes.
2. Pipette and mix 2 μ l of the serum/plasma sample into the tube containing 498 μ l of diluent. This provides a 250 fold diluted sample.
3. Mix 2 μ l of the 250 fold diluted sample with the 798 μ l of diluent in the second tube. This provides a 100,000 fold dilution of the sample.
4. Repeat this procedure for each sample to be tested

Tissue extracts, and tissue culture medium will likely have lower IgG1 levels than those found in serum. Optimal dilutions of such samples should be determined empirically.

ASSAY PROCEDURE

1. Secure the desired number of coated wells in the holder.
2. Dispense 100 μ l of standards and diluted samples into the wells (we recommend that samples be tested in duplicate).
3. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
4. Aspirate the contents of the microtiter wells and wash the wells 5 times with 1x wash solution using a plate washer (400 μ l/well). The entire wash procedure should be performed as quickly as possible.
5. Strike the wells sharply onto absorbent paper or paper towels to remove all residual wash buffer.
6. Add 100 μ l of enzyme conjugate reagent into each well.
7. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 30 minutes.
8. Wash as detailed in 4 to 5 above.
9. Dispense 100 μ l of TMB Reagent into each well.
10. Gently mix on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 20 minutes.
11. Stop the reaction by adding 100 μ l of Stop Solution to each well.
12. Gently mix. *It is important to make sure that all the blue color changes to yellow.*
13. Read the optical density at 450 nm with a microtiter plate reader within 5 minutes.
- 14.

CALCULATION OF RESULTS

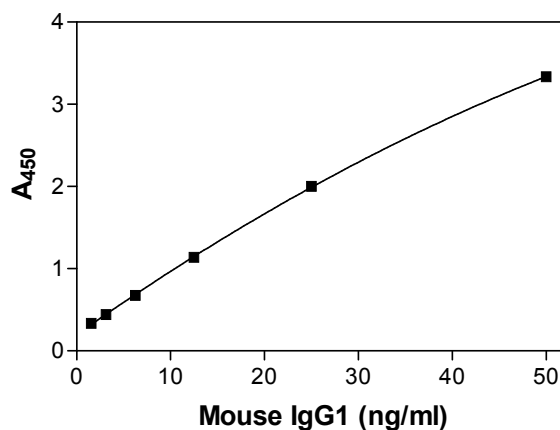
1. Calculate the average absorbance values (A_{450}) for each set of reference standards and samples.
2. Construct a standard curve by plotting the mean absorbance obtained from each reference standard against its concentration in ng/ml on linear graph paper, with absorbance values on the vertical or Y-axis and concentrations on the horizontal or X-axis.
3. Using the mean absorbance value for each sample, determine the corresponding concentration of IgG1 in ng/ml from the standard curve.
4. Multiply the derived concentrations by the dilution factor to determine the actual concentration of IgG1 in the sample.
5. PC graphing software may be used for the above steps.
6. If the OD_{450} values of samples fall outside the standard curve, samples should be diluted appropriately and re-tested.

TYPICAL STANDARD CURVE

A typical standard curve with optical density readings at 450nm on the Y axis against IgG1 concentrations on the X axis is shown below. This curve is for the purpose of illustration only and should not be used to calculate unknowns. Each user should obtain his or her data and standard curve in each experiment.

IgG1 (ng/ml)	Absorbance (450 nm)
50	3.334
25	2.002
12.5	1.139
6.25	0.675
3.13	0.441
1.56	0.336

Typical Mouse IgG1 Standard Curve



LIMITATIONS OF THE PROCEDURE

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of and in accordance with the instructions detailed above.
2. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.